

BIOFERTILIZATION AND ORGANIC MANURING EFFICIENCY ON GROWTH AND YIELD OF POTATO PLANTS (*Solanum tuberosum* L.)

By
R.A. Zaghloul

*Agric Botany Department, Faculty of Agriculture, Moshtohor
Zagazig University Banha Branch, Egypt*

ABSTRACT

The aim of this research was to study the possibility of biofertilizers using and organic manuring (biogas fertilizer) instead of chemical fertilizers for potato growth and production. Two field experiments were carried out at the Vegetables Experimental Farm of Fac. Agric. Moshtohor, Zagazig Univ. during two successive spring seasons of 2001 and 2002. *Azotobacter chroococcum* and *Azospirillum lipoferum* were used as nitrogen fixing bacteria. While, *B. megaterium* var. *phosphaticum* was used as phosphate solubilizing bacteria. Biogas manure and ammonium sulphate were used as organic and inorganic fertilizers, respectively. Biogas manure was added at a rate of 6 ton/fed. (90 kg N/ fed.) as well as using ammonium sulphate at the same level of nitrogen.

Obtained results indicated that the highest records of evolved CO₂ were observed in biogas manure treatments. Whereas, the highest records of N₂-ase activity were observed in rhizosphere of potato plants inoculated with asymbiotic N₂- fixers. Inoculation of potato tubers with phosphate solubilizing bacteria combined with various treatments under investigation increased CO₂ evolution and N₂-ase activity.

Inoculation of potato tubers with *Azotobacter* & *Azospirillum* and a half dose of ammonium sulphate receiving gave higher records of NH₄- N and NO₃ - N in rhizosphere soil than the application of full dose from ammonium sulphate. Biogas manure amendment showed the highest records of N and P in rhizosphere soil.

Growth characteristics of potato plants were significantly increased with biogas manure application in combination with potato tuber inoculation with phosphate solubilizing bacteria. Potato tuber inoculated with *Azotobacter* & *Azospirillum* combined with *B. megaterium* var. *phosphaticum* showed the highest records of carbohydrate content in tubers. Insignificant difference in tuber yield/ fed. was observed between asymbiotic N₂ - fixing bacteria and biogas manuring treatments. Generally, tuber yield/fed. was higher with *Azotobacter* & *Azospirillum* and biogas manuring treatments than ammonium sulphate application. Therefore, the use of biofertilizers or organic manuring may be recommended as a substitute for chemical fertilizers in potato crop production especially for exportation.

Key words: potato, *A. chroococcum*, *A. lipoferum*, *B. megaterium* var. *phosphaticum*, biogas manure.

1. INTRODUCTION

Potato (*Solanum tuberosum* L.) is one of the most important vegetable crops cultivated in Egypt for local consumption and exportation. Increasing the quality of potato for exportation is the main aim of potato growers. Nitrogen and phosphatic chemical fertilizers are commonly used in production of vegetable crops. Application of such chemical fertilizers to the soil causes some problems especially for exportation. It is well known that the nitrogenous fertilizers are lost via nitrate reduction, denitrification and ammonia volatilization. Moreover, some nitrogenous fertilizers can be leached to the surface and underground water causing environmental pollution (Attia, 1990). Also, immobilization of phosphorus is the most important problem of phosphatic fertilization in Egypt and this is due to soil alkalinity. Taking the economical point into account, the high prices of chemical fertilizers may increase the production costs of potato producers.

Therefore, the use of biofertilizers and organic manures is a particular interest to avoid the previously mentioned problems. Biofertilizers application with a half dose of chemical nitrogen fertilizer proved to be an efficient tool in increasing available nutrients in soil as well as growth performance and yield of cultivated crops is improved. Several investigators indicated that inoculation with *Azotobacters* and *Azospirilla* improved growth and yield of potato crop (Mahendran *et al*, 1996; Zahir and Muhammed, 1996; Zahir *et al*, 1997; Jadhav *et al* 1998, Mahendran and Kumar, 1998 and El-Ghinbihi and Fetouh, 2001).

With regard to the effect of organic manuring on plant growth and microbial activity. Abdel- Magid *et al* (1996); Zaghloul *et al* (1996) and Neweigy *et al* (1997) reported that the addition of organic manures to the soil encouraged proliferation of soil microorganisms, increased microbial populations and activity of microbial enzymes i.e Dehydrogenase, Urease and Nitrogenase. In addition, Sood (1993); Sharma (1993); Karadogan (1995); Merghany (1998) and El- Fakhrani (1999) found that organic manuring had a significant stimulative effect on growth characters and tuber yield of potato plants when compared with application of the same level of nitrogen from inorganic nitrogen fertilizers. Concerning the effect of dual inoculation with asymbiotic N₂- fixing bacteria and phosphate solubilizing bacteria on potato growth, Abdel- Ati *et al* (1996); El- Gamal (1996) and Mahendran and kumar (1998) found that dual inoculation of potato with asymbiotic N₂- fixers i.e *Azotobacters* or *Azospirilla* and phosphate solubilizing bacteria improved growth performance, dry matter, carbohydrates content and tuber yield of potato. Also, Mahendran and Chandramani (1998) reported that dual inoculation of potato with the above - mentioned microorganisms increased soil available N,P and K.

Therefore, the present investigation was designed to evaluate the effect of biofertilization and organic manuring on growth and yield of potato.

2. MATERIALS AND METHODS

Two field experiments were carried out during the spring seasons of 2001 and 2002 at the Agricultural Research and Experimentation Center (vegetables farm) of Fac. Agric; Moshtohor, Zagazig Univ. to study the effect of nonsymbiotic N₂- fixing bacteria, biogas manure and ammonium sulphate application either individually or combined with phosphate solubilizing bacteria on growth and yield of potato cv. Diamant. Physical and chemical analyses of the experimental soil are shown in Table (1). Also, the chemical analysis of biogas manure is shown in Table (2).

Table 1. Physical and chemical analyses of the experimental soil.

| Parameters | Unit | Seasons | |
|-------------------------------|------|-----------|-----------|
| | | 2001 | 2002 |
| A. Mechanical analysis | | | |
| Coarse sand | (%) | 3.62 | 3.15 |
| Fine sand | (%) | 23.12 | 25.18 |
| Silt | (%) | 26.50 | 27.12 |
| Clay | (%) | 46.76 | 44.55 |
| Textural class | | Clay loam | Clay loam |
| B. Chemical analysis | | | |
| Organic matter | (%) | 1.52 | 1.67 |
| Total nitrogen | (%) | 0.26 | 0.28 |
| Total phosphorus | (%) | 0.14 | 0.18 |
| Total potassium | (%) | 0.31 | 0.34 |
| pH | | 8.31 | 8.11 |

Table 2. Chemical analysis of biogas manure.

| Parameters | Unit | Seasons | |
|------------------|-------|---------|-------|
| | | 2001 | 2002 |
| Organic matter | (%) | 61.30 | 58.21 |
| Organic carbon | (%) | 32.55 | 33.76 |
| Total nitrogen | (%) | 1.48 | 1.56 |
| Total phosphorus | (%) | 0.81 | 0.86 |
| Total potassium | (%) | 1.20 | 1.28 |
| C:N ratio | | 24.02 | 21.64 |
| Iron | (ppm) | 48.1 | 45.5 |
| Zinc | (ppm) | 36.2 | 32.1 |
| Copper | (ppm) | 23.3 | 27.6 |
| Manganese | (ppm) | 16.5 | 21.3 |

Mechanical analysis was estimated according to Jackson (1973). While, chemical analysis of soil and biogas manure was estimated according to Black *et al* (1982).

2.1. Potato tubers

Certified potato tubers (cv. Diamant) were obtained from the general authority for producers and Exporters of Horticultural Crops, Cairo, Egypt.

2.2. The used microorganisms

Azotobacter chroococcum UF5 and *Azospirillum lipoferum* Mn3 strains were provided from the unit of Biofertilization, Fac. of Agric., Ain Shams Univ., Cairo, Egypt. While, *Bacillus megaterium* var *phosphaticum* (pure local strain) was provided from Microbiology Dept. Soil & Water and Environment Res. Inst., Agric., Res. Center, Giza, Egypt.

2.3. Inocula preparation

For preparation of *A. chroococcum* and *A. lipoferum* inocula, modified Ashby's medium (Abdel-Malek and Ishac, 1968) and semi-solid malate medium (Dobereiner, 1978) were inoculated with *A. chroococcum* and *A. lipoferum*, respectively then incubated at 30°C and 32°C for 7 days, respectively. Also, Bunt and Rovira medium (1955) modified by Abdel-Hafez (1966) was inoculated with *B. megaterium* var. *Phosphaticum* then incubated at 30°C for 7 days.

Biogas manure was added to the soil before sowing at a rate of 6 ton/ fed. (90kg N/fed). Chemical phosphorus fertilizer at a rate of 31kg P₂O₅ in form of calcium superphosphate (15.5% P₂O₅) was applied during preparation of the soil for all treatments. Nitrogen and potassium fertilizers were added at rates of 90 kg N/ fed. and 96 kg K₂O / fed. in forms of ammonium sulphate (20.5% N) and potassium sulphate (48%K₂O) in three equal doses at 15, 30 and 60 days after emergence.

2.4. Inoculation process

Pieces of potato tuber were successfully washed with water and air dried. Thereafter, they were soaked in cell suspension of a mixture (1:1) from *A. chroococcum* and *A. lipoferum* (1ml contains about 8X10⁷ viable cells) for 30 minutes. Tubers of control treatment was treated with the same manner but using N- deficient medium instead of bacterial cultures.

Potato tuber pieces of all treatments were divided into two parts, the first part was planted without inoculation with phosphate solubilizing bacteria whereas, the second part was planted after soaking in cell suspension of *B. megaterium* var. *phosphaticum* (1ml contains about 10⁸ viable cells) for 30 minutes. Sucrose solution (30%) was added as an adhesive agent prior to inoculation.

2.5. Experimental design

Treatments were distributed in a randomized complete block design with three replicates. This experiment included the following treatments:

- 1- Control (without any addition).
- 2- *A.chroococcum* & *A.lipoferum* inoculum (1:1) + a half dose of inorganic N- fertilizer (hdn).
- 3- A full dose of biogas manure, 6 ton/fed. (90 kg N/fed.).

- 4- A full dose of ammonium sulphate (90kgN/fed).
- 5- Control + *B. megaterium* (PSB) inoculum.
- 6- *A. chroococcum* & *A. lipoferum*+ hdn+ PSB.
- 7- A full dose of biogas manure+ PSB.
- 8- A full dose of ammonium sulphate+ PSB.

2.6. Cultivation Process

Cultivation process was performed at January 20th by sowing uninoculated or inoculated tuber pieces in ridges 5m long and 70 cm a part. Other field practices for potato growing were followed according to the recommendations of the Ministry of Agriculture.

2.7. Sampling and determinations

Representative soil samples from rhizosphere of potato plants were taken at 7, 15, 30, 60, and 90 days from sowing. The samples were microbiologically analyzed for densities of *Azotobacters*, *Azospirilla*, phosphate solubilizing bacteria, CO₂ evolution and N₂-ase activity. Also, rhizosphere soil samples were chemically analyzed at 15, 30, 60, and 90 days for ammoniacal and nitrate nitrogen, total nitrogen and phosphorus.

2.7.1.A. Microbiological analysis

A.1. Densities of *Azotobacters* and *Azospirilla* were determined on modified Ashby's medium (Abdel- Malek and Ishac, 1968) and Semi-solid malate medium (Dobereiner, 1978), respectively using the most probable number technique (Cochran, 1950) whereas, the density of inorganic phosphate dissolvers was determined on (Bunt and Rovira medium, 1955 modified by Abdel- Hafez, 1966) using the plate count method.

A.2. Carbon dioxide evolved by soil microorganisms was estimated according to page *et al* (1982).

A.3. Nitrogenase activity was estimated according to Hardy *et al* (1973).

2.7.2.B. Chemical analysis of soil

B.1. Ammoniacal and nitrate nitrogen were determined according to Bremner and Keeny (1965)

B.2. Total nitrogen was estimated according to A.O.A.C (1980). Whereas, total phosphorus was estimated according to A.P.H.A (1992).

2.7.3. Growth Parameters

After 70 days from sowing, plant height, dry matter of shoot system, leaves number/plant and branches number/ plant were estimated.

2.7.4. Chemical analysis of plant

Total nitrogen and phosphorus were periodically determined in dried leaves at 15, 30, 60, and 90 days from sowing.

At harvesting, number of tubers/kg, percentage of total carbohydrate in tubers and tuber yield/fed. were estimated. Total carbohydrate was estimated according to Michel *et al* (1956)

2.8. Statistical analysis

Analysis of variance (ANOVA) of data obtained from growth characters, yield and yield component were carried out according to Snedecor and Cochran (1989). The differences between the means values of various treatments were compared by Duncan's multiple range test (Duncan's, 1955).

3. RESULTS AND DISCUSSION

3.1. Effect of biofertilization and organic manuring on microbial densities in rhizosphere of potato plants.

Data in Table (3) show the periodical changes of *Azotobacters*, *Azospirilla* and phosphate solubilizing bacteria in rhizosphere of potato plants. Microbial densities of the abovementioned bacterial groups gradually increased with the increasing of growth period to reach their maximum records at 60 days and decreased thereafter. The same trend of results was observed in all treatments. Rhizosphere of potato plants inoculated with *Azotobacter* & *Azospirillum* and receiving a half dose of ammonium sulphate contained higher densities of

Azotobacters and *Azospirilla* than either biogas manure or ammonium sulphate application. Such results may indicate that the introduced inoculum has the ability to survive and colonize the root zone of potato plants. Similar results were obtained by **Abdel-Ati et al (1996)**; **Shatokhina and Khristenko (1996)** and **Saleh et al (1998)**. Regarding the densities of phosphate solubilizing bacteria, obtained data show that rhizosphere of potato plants amended with biogas manure contained the highest densities of p-solubilizing bacteria compared to other treatments. Irrespective of control, the lowest densities of p-solubilizing bacteria was observed in the treatment of ammonium sulphate application. The same trend of results was observed at various determination periods. This result is in accordance with **Abdel-Magid et al (1996)** and **Neweigy et al (1997)**.

It is worthy to notice that the rhizosphere of potato plants inoculated with phosphate solubilizing bacteria contained higher densities of *Azotobacters*, *Azospirilla* and phosphate dissolvers compared to the uninoculated ones. This was true in all treatments and different determination periods. These results are in harmony with **Zaghloul et al (1996)** and **Saad and Hammad (1998)**.

3.2. Effect of biofertilization and organic manuring on CO₂ evolution and N₂-ase activity in rhizosphere of potato plants.

Data presented in Table (4) indicate that carbon dioxide evolved by soil microorganisms gradually increased throughout the experimental period to reach their maximum values at 60 days and decreased thereafter. The same trend of results was obtained in both growing seasons and various investigated treatments. The highest records of evolved carbon dioxide was observed in biogas manure treatment compared to other treatments when applied alone. As well, this trend of results was obtained in case of biogas manure application combined with phosphate solubilizing bacteria inoculum. Rhizosphere of potato plants inoculated with *Azotobacter* & *Azospirillum* showed higher values of carbon dioxide evolution than either uninoculated one or ammonium sulphate application. Inoculated treatments with phosphate solubilizing bacteria showed higher values of CO₂ evolution than uninoculated treatments. This was observed in both growing seasons as well at all determination periods. This result may explain the role of *B.megaterium* var. *phosphaticum* in supplying different soil microorganisms and plants with their available phosphorus requirements and consequently increase the activity of soil microorganisms. It is clear from data presented in Table (4) that the highest records of N₂-ase activity were recorded in the rhizosphere of potato plants inoculated with asymbiotic N₂-fixers and supplemented with a half dose of inorganic N-fertilizer. Also, rhizosphere of potato plants supplemented with biogas manure showed higher records of N₂-ase activity than the application of full dose from ammonium sulphate. These results are in agreement with **Mahendran et al (1996)**; **Zahir et al (1997)** and **Saleh et al (1998)**. They reported that biofertilization of plants with asymbiotic N₂-fixers increased N₂-ase activity compared to unbiofertilized ones. Also, they found that increasing the dose of inorganic N-fertilizer resulted a decreasing in N₂-ase activity. Moreover, it could be concluded that inorganic N-fertilization exhibited a negative effect on biological N₂ fixation. Nevertheless, low doses of N-fertilizer promoted the response of cultivated plants to inoculation with asymbiotic N₂-fixers (**El-Demerdash, 1994** and **Jadhav et al 1998**). They reported that half of the recommended dose of added inorganic N-fertilizer can be saved by asymbiotic N₂-fixers inoculation. Such trends support the obtained results in the current study.

It is obvious from data recorded in Table (4) that when tuber of potato plants inoculated with phosphate solubilizing bacteria, the N₂-ase activity values were increased compared to uninoculated ones. Generally, carbon dioxide evolution and N₂-ase activity values were higher during the second season than the first one. These differences between the two seasons are likely to be due to the differences in the climatic conditions.

Table 3. Azotobacters, Azospirilla and inorganic phosphate solubilizing bacteria densities (number/g dry weight of soil) in rhizosphere of potato plants. (The recorded densities represent the average of the two growing seasons).

| Treatments | Days after sowing | | | | | | | | | |
|----------------------|---|-------|-------|-------|-------|---|-------|-------|-------|-------|
| | 7 | 15 | 30 | 60 | 90 | 7 | 15 | 30 | 60 | 90 |
| | Non inoculated with phosphate solubilizing bacteria | | | | | Inoculated with phosphate solubilizing bacteria | | | | |
| | Azotobacters $\times 10^5$ | | | | | | | | | |
| Control | 10.30 | 24.50 | 45.30 | 48.20 | 31.30 | 22.20 | 36.40 | 52.10 | 62.30 | 36.30 |
| Azoto. & Azos. + hdn | 34.20 | 81.40 | 110.1 | 140.0 | 95.40 | 42.30 | 86.30 | 120.2 | 186.4 | 160.3 |
| Biogas manure | 18.20 | 62.60 | 74.20 | 90.10 | 43.20 | 38.80 | 80.20 | 88.30 | 96.60 | 78.60 |
| Ammonium sulphate | 12.01 | 40.40 | 53.20 | 82.02 | 38.40 | 36.20 | 44.10 | 56.50 | 82.20 | 70.40 |
| | Azospirilla $\times 10^4$ | | | | | | | | | |
| Control | 8.40 | 23.40 | 38.10 | 41.20 | 26.20 | 24.10 | 36.50 | 52.20 | 60.40 | 43.30 |
| Azoto. & Azos. + hdn | 40.20 | 78.60 | 92.10 | 120.1 | 82.10 | 60.20 | 71.30 | 86.30 | 98.30 | 81.40 |
| Biogas manure | 36.20 | 61.30 | 70.20 | 90.60 | 63.40 | 56.30 | 68.20 | 80.20 | 112.1 | 86.40 |
| Ammonium sulphate | 25.50 | 42.20 | 62.10 | 81.30 | 52.20 | 43.20 | 56.60 | 78.10 | 91.10 | 73.40 |
| | phosphate solubilizing bacteria $\times 10^5$ | | | | | | | | | |
| Control | 12.60 | 20.40 | 28.40 | 42.20 | 38.10 | 18.70 | 32.40 | 40.50 | 54.00 | 42.50 |
| Azoto. & Azos. + hdn | 16.40 | 40.20 | 48.40 | 68.00 | 62.30 | 28.20 | 72.20 | 90.30 | 116.2 | 72.30 |
| Biogas manure | 20.20 | 48.60 | 62.00 | 90.30 | 80.00 | 32.40 | 96.30 | 110.2 | 128.0 | 91.40 |
| Ammonium sulphate | 14.40 | 28.20 | 40.20 | 72.40 | 60.30 | 21.10 | 36.30 | 56.00 | 81.30 | 56.60 |

Azoto., Azotobacter.
Azos., Azospirillum.

hdn., half dose of nitrogen.

Table 4. Carbon dioxide evolution and nitrogenase activity in rhizosphere of Potato plants during the two growing seasons.

| Treatments | Days after sowing | | | | | | | | | | | | | | | | | | | |
|--|--|------|-------|-------|------|------|--------|-------|-------|-------|--|------|------|------|------|------|-------|-------|------|------|
| | 7 | | | | 15 | | | | 30 | | | | 60 | | | | 90 | | | |
| | S1 | | S2 | | S1 | | S2 | | S1 | | S2 | | S1 | | S2 | | S1 | | S2 | |
| | CO ₂ evolved (µg/g dry soil/ hr.) | | | | | | | | | | N ₂ -ase activity (n moles C ₂ H ₄ /g dry soil/hr.) | | | | | | | | | |
| Non inoculated with phosphate solubilizing bacteria | | | | | | | | | | | | | | | | | | | | |
| Control | 8.16 | 10.2 | 14.02 | 18.60 | 21.2 | 23.3 | 48.01 | 50.10 | 30.12 | 32.1 | 12.2 | 14.1 | 18.2 | 16.3 | 23.1 | 26.4 | 28.5 | 31.10 | 21.4 | 26.3 |
| Azoto. & Azos. + hdn | 15.1 | 18.2 | 28.64 | 32.30 | 48.1 | 52.2 | 65.20 | 68.10 | 36.80 | 40.3 | 28.3 | 30.6 | 36.2 | 40.1 | 61.2 | 72.1 | 98.3 | 105.1 | 72.1 | 74.2 |
| Biogas manure | 22.1 | 26.4 | 49.29 | 38.30 | 62.6 | 70.3 | 103.10 | 109.1 | 82.40 | 86.8 | 24.6 | 25.5 | 30.4 | 33.1 | 48.2 | 51.1 | 69.2 | 73.10 | 54.3 | 58.5 |
| Ammonium sulphate | 12.6 | 16.1 | 22.91 | 26.40 | 51.4 | 58.2 | 58.20 | 62.60 | 61.30 | 65.2 | 20.3 | 18.6 | 34.4 | 36.2 | 46.1 | 44.3 | 61.4 | 65.20 | 42.1 | 45.1 |
| Inoculated with phosphate solubilizing bacteria | | | | | | | | | | | | | | | | | | | | |
| Control | 11.2 | 13.3 | 18.16 | 20.3 | 31.2 | 36.1 | 51.30 | 56.40 | 46.60 | 42.10 | 22.6 | 25.5 | 26.1 | 30.2 | 43.2 | 46.1 | 48.50 | 51.30 | 30.4 | 36.2 |
| Azoto.& Azos. + hdn | 18.4 | 21.2 | 45.83 | 43.2 | 68.1 | 76.3 | 106.1 | 110.2 | 81.40 | 86.20 | 36.5 | 41.6 | 52.5 | 63.3 | 82.2 | 96.1 | 148.1 | 161.2 | 89.4 | 45.6 |
| Biogas manure | 26.1 | 32.4 | 51.56 | 56.2 | 82.3 | 93.1 | 153.1 | 161.2 | 106.4 | 116.2 | 32.4 | 35.2 | 41.2 | 46.3 | 63.1 | 65.5 | 121.1 | 130.3 | 81.4 | 88.5 |
| Ammonium sulphate | 17.5 | 23.6 | 32.08 | 40.2 | 57.1 | 62.3 | 70.12 | 82.40 | 63.40 | 68.30 | 28.3 | 32.4 | 36.3 | 38.2 | 52.3 | 56.3 | 82.40 | 85.30 | 63.2 | 66.5 |

Abbreviations: as those stated in Table (3)•

S1: The first season.

S2: The second season

3.3. Effect of biofertilization and organic manuring on nitrogen forms in rhizosphere of potato plants.

Data recorded in Table (5) show the periodical changes of $\text{NH}_4\text{-N}$ and $\text{NO}_3\text{-N}$ in rhizosphere of potato plants. Obtained data show that ammoniacal and nitrate nitrogen gradually increased with the progression of growth period and reaching their maximum records at 60 days. This was true in all applied treatments. Biogas manure application increased ammoniacal and nitrate nitrogen in rhizosphere of potato plants.

Inoculation of potato tuber at sowing with *Azotobacter* & *Azospirillum* and a half dose of ammonium sulphate supplementation gave higher records of $\text{NH}_4\text{-N}$ and $\text{NO}_3\text{-N}$ than the application of full dose from ammonium sulphate. Similar results were obtained by **Zaghoul et al (1996)** and **Neweigy et al (1997)** who reported that ammoniacal and nitrate nitrogen content were higher in case of biofertilization with asymbiotic N_2 -fixers than inorganic N-fertilization in rhizosphere soil of wheat and sorghum, respectively.

From data presented in Table (5) it is worthy to mention that the inoculation of potato tuber with *B.megaterium var. phosphaticum* in combination with various investigated treatments increased $\text{NH}_4\text{-N}$ and $\text{NO}_3\text{-N}$ values in comparison with uninoculated ones. This trend of results was observed in both growing seasons and various determination periods. This result may explain the role of p-dissolvers in availability of phosphorus and micro-elements and consequently different soil microbial activity were increased.

3.4. Effect of biofertilization and organic manuring on total nitrogen and phosphorus in rhizosphere of potato plants.

Data in Table (6) indicate that total nitrogen and phosphorus percentages in rhizosphere of potato plants gradually decreased with the progression of growth period and reached their minimum records at 60 days and increased thereafter. The decrease of nitrogen and phosphorus content of soil could be attributed to nutrients uptake by potato plants especially at early growth periods. Rhizosphere soil of potato plants amended with biogas manure showed the highest nitrogen and phosphorus content and this result was observed at all estimation periods.

In general biofertilization with *Azotobacter* & *Azospirillum* showed higher values of nitrogen and phosphorus percentages than fertilization with ammonium sulphate. These results are in agreement with the findings of **Mahendran et al (1996)**; **Zahir et al (1997)** and **Mahendran and Kumar (1998)** who reported that biofertilizers application increase nutrients content in soil in comparison with inorganic N-fertilizers.

Also, data recorded in Table (6) emphasize that inoculation of potato tuber with *B.megaterium var. phosphaticum* in combination with asymbiotic N_2 -fixers, biogas manure and ammonium sulphate increased nutrients content (N and P) in soil compared to uninoculated treatments. Such results may explain the synergistic effect of phosphate solubilizing bacteria. This result is in accordance with **El-Gamal (1996)**; **Mahendran and Chandramani (1998)**. They reported that dual inoculation of potato tuber with *Azospirillum* and phosphate solubilizing bacteria increased soil N and P availability as well as improved potato yield and tuber quality.

3.5. Effect of biofertilization and organic manuring on some growth characters of potato plants.

It is obvious from data in Table (7) that growth characteristics i.e plant height, leaves number/plant and branches number/plant were significantly increased with biogas manure application in combination with potato tuber inoculated with *B.megaterium var. phosphaticum*. This finding was observed in both growing seasons. The high records of potato growth performance which were observed in biogas manure treatments can be explained by their high densities of phosphate solubilizing bacteria (Table, 3) and the increase of ammoniacal and nitrate nitrogen content in rhizosphere soil (Table, 5). Similar results were observed by **Merghany (1998)** and **El-Fakhrani (1999)**. They found that organic manuring had a significant stimulative effect on growth characters of potato plants when compared with application of the same level of nitrogen from inorganic N-fertilizers.

Table 5. Ammoniacal and nitrate nitrogen concentration in rhizosphere of potato plants during the two growing seasons.

| Treatments | Days after sowing | | | | | | | | | | | | | | | |
|----------------------|--|------|------|------|------|------|------|------|------------------------|------|------|------|------|------|------|------|
| | Ammoniacal nitrogen (ppm) | | | | | | | | Nitrate nitrogen (ppm) | | | | | | | |
| | 15 | | 30 | | 60 | | 90 | | 15 | | 30 | | 60 | | 90 | |
| | S1 | S2 | S1 | S2 | S1 | S2 | S1 | S2 | S1 | S2 | S1 | S2 | S1 | S2 | S1 | S2 |
| | Non inoculated with phosphate solubilizing bacteria | | | | | | | | | | | | | | | |
| Control | 8.60 | 9.2 | 14.6 | 18.1 | 26.2 | 30.1 | 16.4 | 14.6 | 9.80 | 10.2 | 13.2 | 15.2 | 18.3 | 20.1 | 12.4 | 14.1 |
| Azoto. & Azos. + hdn | 21.3 | 28.1 | 41.0 | 44.0 | 48.0 | 52.0 | 30.0 | 28.0 | 11.4 | 13.8 | 19.0 | 23.0 | 36.0 | 38.0 | 21.0 | 24.0 |
| Biogas manure | 29.6 | 31.2 | 46.0 | 43.0 | 63.0 | 65.0 | 36.0 | 33.0 | 19.6 | 21.2 | 30.0 | 28.0 | 41.0 | 44.0 | 20.0 | 22.0 |
| Ammonium sulphate | 14.6 | 16.2 | 28.0 | 30.0 | 41.0 | 43.0 | 26.0 | 24.0 | 15.1 | 22.1 | 26.0 | 24.0 | 35.0 | 32.0 | 18.0 | 21.0 |
| | Inoculated with phosphate solubilizing bacteria | | | | | | | | | | | | | | | |
| Control | 12.6 | 14.0 | 19.0 | 21.0 | 32.0 | 34.0 | 20.0 | 18.0 | 12.6 | 14.0 | 16.0 | 18.0 | 21.0 | 23.0 | 15.1 | 14.5 |
| Azoto. & Azos. + hdn | 28.0 | 32.1 | 48.0 | 51.0 | 58.0 | 62.0 | 38.0 | 36.0 | 15.6 | 16.8 | 24.0 | 28.0 | 43.0 | 46.0 | 30.0 | 32.0 |
| Biogas manure | 33.1 | 36.3 | 50.0 | 53.0 | 61.0 | 66.0 | 40.0 | 42.0 | 23.1 | 30.0 | 36.0 | 32.0 | 52.0 | 56.0 | 29.0 | 33.0 |
| Ammonium sulphate | 16.8 | 18.2 | 32.0 | 34.0 | 46.0 | 49.0 | 26.0 | 28.2 | 21.3 | 26.5 | 32.0 | 28.0 | 39.0 | 36.0 | 27.2 | 29.1 |

Abbreviations: as those stated in Table (3)•

S1: The first Season

S2: The second season

Table 6. Periodical changes in total nitrogen and phosphorus in rhizosphere of potato plants during the two growing seasons.

| Treatments | Days after sowing | | | | | | | | | | | | | | | |
|----------------------|---|-------|-------|-------|-------|-------|-------|-------|----------------------|-------|-------|-------|-------|-------|-------|-------|
| | 15 | | 30 | | 60 | | 90 | | 15 | | 30 | | 60 | | 90 | |
| | Total nitrogen (%) | | | | | | | | Total phosphorus (%) | | | | | | | |
| | S1 | S2 | S1 | S2 | S1 | S2 | S1 | S2 | S1 | S2 | S1 | S2 | S1 | S2 | S1 | S2 |
| | Non inoculated with phosphate solubilizing bacteria | | | | | | | | | | | | | | | |
| Control | 0.200 | 0.229 | 0.210 | 0.214 | 0.175 | 0.181 | 0.190 | 0.181 | 0.110 | 0.116 | 0.101 | 0.112 | 0.074 | 0.091 | 0.102 | 0.118 |
| Azoto. & Azos. + hdn | 0.305 | 0.326 | 0.215 | 0.226 | 0.140 | 0.151 | 0.175 | 0.196 | 0.182 | 0.193 | 0.144 | 0.156 | 0.125 | 0.138 | 0.179 | 0.186 |
| Biogas manure | 0.375 | 0.381 | 0.235 | 0.240 | 0.105 | 0.118 | 0.198 | 0.224 | 0.233 | 0.240 | 0.175 | 0.173 | 0.161 | 0.165 | 0.200 | 0.228 |
| Ammonium sulphate | 0.245 | 0.250 | 0.230 | 0.226 | 0.110 | 0.126 | 0.165 | 0.188 | 0.144 | 0.152 | 0.126 | 0.131 | 0.122 | 0.118 | 0.135 | 0.142 |
| | Inoculated with phosphate solubilizing bacteria | | | | | | | | | | | | | | | |
| Control | 0.232 | 0.238 | 0.220 | 0.196 | 0.116 | 0.121 | 0.165 | 0.178 | 0.130 | 0.136 | 0.122 | 0.128 | 0.096 | 0.110 | 0.118 | 0.122 |
| Azoto. & Azos. + hdn | 0.355 | 0.362 | 0.280 | 0.261 | 0.158 | 0.163 | 0.211 | 0.236 | 0.173 | 0.182 | 0.153 | 0.161 | 0.141 | 0.148 | 0.172 | 0.180 |
| Biogas manure | 0.418 | 0.426 | 0.312 | 0.260 | 0.187 | 0.168 | 0.242 | 0.286 | 0.337 | 0.361 | 0.184 | 0.190 | 0.153 | 0.163 | 0.230 | 0.242 |
| Ammonium sulphate | 0.265 | 0.271 | 0.250 | 0.231 | 0.155 | 0.172 | 0.195 | 0.220 | 0.153 | 0.171 | 0.136 | 0.143 | 0.126 | 0.121 | 0.145 | 0.138 |

Abbreviations: as those stated in Table (3) •

The records of growth parameters were higher in the asymbiotic N₂-fixers treatments than ammonium sulphate application. This may be due to the production of growth regulators such as auxins, cytokinins and gibberellins by asymbiotic N₂-fixing bacteria which affect the production of root biomass and nutrients uptake (Fulchieri and Frioni, 1994; Jadhav *et al* 1998 and El-Ghinbihi and Fetouh, 2001). The same trend of results was obtained in both growing seasons.

In general, growth parameters of potato plants were higher in all investigated treatments which inoculated with phosphate solubilizing bacteria compared to uninoculated ones.

3.6. Effect of biofertilization and organic manuring on total nitrogen and phosphorus.

Data in Table (8) show the periodical changes in nitrogen and phosphorus content in potato plants. Obtained data show that total nitrogen and phosphorus content was increased with the increasing of growth period and reached their maximum values at 60 days. The increase of nitrogen and phosphorus content was correspondent with the decrease of nitrogen and phosphorus content in rhizosphere soil of potato plants (Table, 6). Inoculation of potato tubers with asymbiotic N₂-fixing bacteria gave higher records of total nitrogen in potato plants compared to either biogas manure or ammonium sulphate application. The highest records of nitrogen content was observed in asymbiotic N₂-fixers inoculated treatments in combination with *B.megaterium var. phosphaticum*. El-Gamal (1996) and Mahendran and Chandramani (1998) found that dual inoculation of potato tubers with *Azospirillum* and phosphate solubilizing bacteria increased N, P and K content of potato plants. Biogas manure application either solely or in combination with phosphate solubilizing bacteria showed higher records of total phosphorus in comparison with other investigation treatments. Also, biogas manure treatments gave higher values of nitrogen and phosphorus content compared to ammonium sulphate application. The increase of such nutrients content in biogas manuring can be attributed to the high records of *Azotobacters*, *Azospirilla* and phosphate solubilizing bacteria densities in biogas manure treatment compared to ammonium sulphate application which previously discussed in Table (3). These results are in harmony with Sharma (1993); Karadogan (1995) and Merghany (1998).

Generally, inoculation of potato tubers with phosphate solubilizing bacteria in combination with various investigation treatments improved nitrogen and phosphorus content in potato plants. This was observed at all determination periods as well as in both growing seasons.

3.7 Effect of biofertilization and organic manuring on dry matter, tuber numbers per kg, carbohydrate content and tuber yield.

Data in Table (9) indicate that dry matter per plant was significantly increased in biogas manure application in combination with *B.megaterium var. phosphaticum* inoculum. *Azotobacter* & *Azospirillum* inoculum in combination with phosphate solubilizing bacteria significantly decreased the number of tubers/kg. It is worthy to mention that the decrease of tubers number /kg is desirable character for consumers.

Irrespective of control, the highest number of tubers/kg was observed with ammonium sulphate application.

Respecting the effect of different investigated treatments on carbohydrate content in potato tubers, obtained data show that potato tuber inoculated with *Azotobacter* & *Azospirillum* combined with *B.megaterium var. phosphaticum* showed the highest records of carbohydrate content. Biogas manure application showed higher values of carbohydrate content than ammonium sulphate application. This result was consistent in case of biogas manure application either alone or in combination with phosphate solubilizing bacteria. These results are confirmed with those obtained by Merghany (1998) and El-Fakhrani (1999) who found that the organic manuring significantly increased the total amount of carbohydrates in dry matter of potato tubers in comparison with inorganic N-fertilization.

Data in Table (9) also show that dual inoculation of potato tubers with *Azotobacter* & *Azospirillum* and phosphate solubilizing bacteria significantly increased tuber yield/fed. compared to other investigated treatments. The increase of tubers yield/fed. of potato plants due

Table 7. Effect of biofertilization and organic manuring on some growth characters of Potato plants.

| Growth characters Treatments | Plant height (cm) | | Leaves number/plant | | Branches number / plant | |
|---|-------------------|-----------------|---------------------|-------------------|-------------------------|------------------|
| | S1 | S2 | S1 | S2 | S1 | S2 |
| Non inoculated with phosphate solubilizing bacteria | | | | | | |
| Control | 38 ^h | 41 ^h | 8.3 ^f | 8.60 ^f | 1.6 ^d | 1.3 ^d |
| Azoto.&Azos+hdn | 42 ^f | 45 ^f | 12.6 ^c | 11.6 ^d | 3.3 ^b | 3.0 ^b |
| Biogas manure | 66 ^d | 55 ^c | 16.6 ^a | 16.3 ^a | 3.6 ^b | 3.3 ^b |
| Ammonium sulphate | 40 ^g | 43 ^g | 11.9 ^d | 10.6 ^e | 2.6 ^c | 2.3 ^c |
| Inoculated with phosphate solubilizing bacteria | | | | | | |
| Control | 58 ^c | 56 ^d | 10.3 ^b | 10.6 ^c | 3.0 ^b | 3.0 ^b |
| Azoto.&Azos+hdn | 78 ^b | 79 ^b | 14.6 ^a | 16.3 ^b | 4.3 ^a | 4.6 ^a |
| Biogas manure | 80 ^a | 81 ^a | 16.3 ^a | 16.6 ^b | 4.3 ^a | 4.6 ^a |
| Ammonium sulphate | 70 ^c | 71 ^c | 12.6 ^b | 14.3 ^c | 4.6 ^a | 3.6 ^b |

Abbreviations: as those stated in Table (3).

Means followed by the same letter (s) within each column are not significantly from each other at 5% level.

Table 8. Periodical changes in total nitrogen and phosphorus content of potato plants during the two growing seasons.

| Treatments | Days after sowing | | | | | | | | | | | | | | | |
|---|--------------------|------|------|------|------|------|------|------|----------------------|-------|-------|-------|-------|-------|-------|-------|
| | 15 | | 30 | | 60 | | 90 | | 15 | | 30 | | 60 | | 90 | |
| | Total nitrogen (%) | | | | | | | | Total phosphorus (%) | | | | | | | |
| | S1 | S2 | S1 | S2 | S1 | S2 | S1 | S2 | S1 | S2 | S1 | S2 | S1 | S2 | S1 | S2 |
| Non inoculated with phosphate solubilizing bacteria | | | | | | | | | | | | | | | | |
| Control | 0.86 | 0.92 | 1.58 | 1.62 | 2.32 | 2.40 | 1.74 | 1.83 | 0.109 | 0.118 | 0.142 | 0.162 | 0.210 | 0.228 | 0.192 | 0.210 |
| Azoto. & Azos. + hdn | 1.68 | 1.63 | 2.40 | 2.61 | 3.42 | 3.70 | 2.92 | 2.86 | 0.212 | 0.226 | 0.362 | 0.381 | 0.465 | 0.510 | 0.338 | 0.425 |
| Biogas manure | 1.34 | 1.43 | 1.98 | 2.24 | 3.75 | 3.95 | 2.30 | 2.51 | 0.263 | 0.281 | 0.34 | 0.362 | 0.627 | 0.591 | 0.421 | 0.434 |
| Ammonium sulphate | 0.98 | 1.06 | 1.82 | 1.95 | 3.20 | 3.48 | 1.93 | 2.12 | 0.186 | 0.218 | 0.276 | 0.286 | 0.380 | 0.376 | 0.250 | 0.318 |
| Inoculated with phosphate solubilizing bacteria | | | | | | | | | | | | | | | | |
| Control | 0.84 | 0.96 | 1.62 | 1.70 | 2.86 | 3.12 | 2.21 | 2.45 | 0.141 | 0.153 | 0.186 | 0.192 | 0.240 | 0.228 | 0.200 | 0.224 |
| Azoto. & Azos. + hdn | 1.96 | 1.81 | 2.54 | 2.48 | 4.10 | 4.25 | 3.60 | 3.86 | 0.290 | 0.320 | 0.381 | 0.416 | 0.602 | 0.590 | 0.481 | 0.510 |
| Biogas manure | 1.83 | 1.62 | 2.12 | 2.61 | 3.88 | 4.12 | 3.24 | 3.42 | 0.311 | 0.332 | 0.406 | 0.431 | 0.681 | 0.611 | 0.492 | 0.510 |
| Ammonium sulphate | 1.12 | 1.26 | 1.96 | 2.26 | 3.45 | 3.72 | 2.83 | 2.96 | 0.196 | 0.236 | 0.335 | 0.361 | 0.568 | 0.530 | 0.391 | 0.428 |

Abbreviations: as those stated in Table (3) •

to inoculation with asymbiotic N₂-fixer (*Azotobacters & Azospirillum*) could be attributed to the capability of these organisms to fix nitrogen which could be taken by the growing plants. Similar results were observed by Mahendran *et al* (1996), Mahendran and Kumar (1998) and El-Ghinbihi and Fetouh (2001) who reported that potato tubers inoculated with asymbiotic N₂-fixers in combination with *B.megaterium var. phosphaticum* gave the highest tubers yield /fed.

Data in Table (9) also show that insignificant difference in tuber yield/fed. was observed with biogas manuring amendment compared to biofertilization treatment.

Irrespective of control treatment, the lowest tuber yield/fed. was obtained with ammonium sulphate application.

Generally, from data presented in Table (9) it can be concluded that the inoculation of potato tubers with phosphate solubilizing bacteria in combination with either asymbiotic N₂-fixers or organic manuring as well as inorganic nitrogen fertilization improved tuber yield /fed. of potato plants.

Table 9. Effect of biofertilization and organic manuring on the dry matter, tuber number / kg, carbohydrate content and tuber yield.

| Parameters | Dray matter g / plant | | Tuber number/kg | | Carbohydrate content (%) | | Tuber yield Ton/fed | |
|----------------------|---|-------------------|--------------------|--------------------|-----------------------------|-------------------|------------------------|--------------------|
| | S1 | S2 | S1 | S2 | S1 | S2 | S1 | S2 |
| | Non inoculated with phosphate solubilizing bacteria | | | | | | | |
| Control | 8.50 ^f | 9.00 ^h | 12.66 ^a | 12.63 ^a | 28.1 ^h | 30.6 ^h | 5.80 ^f | 6.20 ^c |
| Azoto. & Azos. + hdn | 28.2 ^c | 26.0 ^c | 8.42 ^d | 8.33 ^d | 50.2 ^d | 52.6 ^d | 10.0 ^c | 12.0 ^{ab} |
| Biogas manure | 33.3 ^b | 36.0 ^b | 8.66 ^d | 8.60 ^d | 44.8 ^c | 46.9 ^c | 10.5 ^c | 9.10 ^c |
| Ammonium sulphate | 16.1 ^d | 22.0 ^f | 10.60 ^c | 10.33 ^c | 39.4 ^f | 41.3 ^f | 8.90 ^d | 11.2 ^b |
| | Inoculated with phosphate solubilizing bacteria | | | | | | | |
| Control | 13.0 ^c | 13.5 ^h | 11.33 ^b | 11.66 ^b | 33.3 ^g | 31.5 ^g | 6.40 ^c | 6.80 ^d |
| Azoto. & Azos. + hdn | 36.0 ^a | 34.0 ^c | 7.83 ^e | 7.21 ^e | 62.4 ^a | 63.1 ^a | 12.9 ^a | 13.4 ^a |
| Biogas manure | 37.0 ^a | 38.0 ^a | 9.33 ^d | 8.90 ^d | 60.2 ^b | 61.8 ^b | 12.2 ^a | 12.9 ^{ab} |
| Ammonium sulphate | 35.0 ^{ab} | 31.6 ^b | 10.33 ^c | 10.61 ^c | 52.4 ^c | 53.3 ^c | 11.2 ^b | 12.0 ^{ab} |

Abbreviations: as those stated in Table (3).

Means followed by the same letter (s) within each column are not significantly different from each other at 5% level.

CONCLUSION

From the obtained results, it can be concluded that potato tuber inoculation with asymbiotic N₂-fixers (*Azotobacter & Azospirillum*) and a half dose of inorganic N-fertilizer supplementation improved growth characters, carbohydrate content and consequently gave higher tuber yield/fed. than using full dose of inorganic N fertilizer. About 50% of the cost of inorganic N-fertilizer can be saved with asymbiotic N₂-fixers inoculation.

Moreover, it is preferable now to use organic manuring and biofertilizers in cultivation. Also, the growth characteristics, carbohydrate content and yield of potato plants inoculated with asymbiotic N₂-fixers combined with phosphate solubilizing bacteria were almostly corresponding to those fertilized with biogas manure combined with phosphate solubilizing bacteria. Therefore, the use of biofertilizers or organic manuring may be recommended as a substitute for chemical fertilization to improve potato productivity and quality to face the local consumption and exportation.

4. REFERENCES

- Abdel-Ati, Y. Y.; Hammad, A. M. A. and Ali, M. Z. H. (1996). Nitrogen fixing and phosphate solubilizing bacteria as biofertilizers for potato plants under Minia conditions. 1st Egyptian Hungarian Horticultural Conf. Kafr El-Sheikh, Egypt. Vol. (1): 25-34.
- Abdel-Hafez, A. M. (1966). Some studies on acid producing microorganisms in soil and rhizosphere with special reference to phosphate dissolvers. Ph. D. Thesis, Agric. Botany Dept. Fac. of Agric. Ain Sahms Univ., Egypt.

- Abdel-Magid, H. M.; Rabie, R. K.; Sabrah, R. E. A. and Abdel -Aal, Sh. I. (1996). The interrelationship between microbial numbers, application rate and biodegradation in a sandy soil. *Arab Gulf Journal Scient. Res.*, 14 (3): 641-657.
- Abd-El-Malek, Y. and Ishac, Y. Z. (1968). Evaluation of methods used in counting of *Azotobacter*. *Journal of Appl. Bact.*, 31: 267-275.
- A. O. A. C. Association of Official Agricultural Chemists (1980). Official Methods of Analysis. 10th Ed. Washington, DC, U. S. A. P: 832.
- A. P. H. A., American Public Health Association (1992). Standard methods for the examination of water and waste water. Washington, Dc, U. S. A.
- Attia, M. A. (1990). The biochemistry of urea decomposition by soil organisms. Ph. D. Thesis. Fac. Agric. Minia Univ., Egypt.
- Bermner, J. M. and Keeny, D. R (1965). Steam distillation method for determination of ammonium, nitrate and nitrite. *Annals Chem. Acta*, 32: 485-495.
- Black, C. A.; Evans, D. O.; Ensminger, L. E.; White, J. L.; Clark, F. E. and Dinauer, R. C. (1982). Methods of Soil Analysis. Part 2. Chemical and microbiological properties. 2nd Ed. Soil Sci., Soc. of Am. Inc. Publ., Madison, Wisconsin, U. S. A.
- Bunt, J. S. and Rovira, A. D. (1955). Microbiological studies of some subarctic soils. *Journal of Soil Sci.*, 6: 114-128.
- Cochran, W. G. (1950). Evaluation of bacterial densities by means of the "most probable densities" *Biometrics*, 6: 105-116.
- Dobereiner, J. (1978). Influence of environmental factors on the occurrence of *S.lipoferum* in soil and roots. *Ecol. Bull.*, Stockholm 26:343-352.
- Duncan's, D. B. (1955). Multiple range and multiple F.test. *Biometrics*, II: 11-24.
- El-Demerdash, M. E. (1994). Growth and yield of maize plants as affected by bacterization with diazotrophs and supplementation with half the dose of N-fertilizer. *Annals of Agric. Sci.*, Moshtohor, 32 (2): 889-898.
- El-Fakhrani, Y. M. (1999). Response of potato plants irrigated with different levels of saline water to organic manuring and N-fertilization. *Annals of Agric. Sci.*, Moshtohor, 37 (2): 1553-1564.
- El-Gamal, A. M. (1996). Response of potatoes to phosphorus fertilizer levels and phosphorous biofertilizer in the newly reclaimed areas. *Assiut Journal of Agric. Sciences*, 27 (2): 77-87.
- El-Ghinbihi, Fatma, H. and Fetouh, A.A. (2001). Response of some potato cultivars to biofertilizers and different mineral nitrogen levels. *Zagazig J. Agric., Res. Vol.* (28) No. (1): 133-162.
- Fulchieri, M. and Frioni, L. (1994). *Azospirillum* inoculation on maize: Effect on yield in a field experimental in central Argentina. *Soil Biology and Biochemistry*, 26 (7): 421-423.
- Hardy, R. W. F.; Bumc, B. C. and Holsten, R. D. (1973). Application of the acetylene-ethylene assay for measurement of nitrogen fixation. *Soil Biol. Biochem.*, 5: 47-81.
- Jackson, M. L. (1973). Soil Chemical Analysis. Prentice-Hall of India private New Delhi.
- Jadhav, A. C.; Memane, S. A. and Konde, B. K. (1998). Evaluation of biofertilizers in respect to growth and yield attributing parameters in sweet potato. *Journal of Maharashtra Agricultural Universities*, 23 (1): 80-82.
- Karadogan, T. (1995). Effects of manures and fertilizer application on yield, yield components and quality of potatoes. *Turkish Journal of Agriculture and Forestry*, 19 (5): 373-377 (field crop Abstr. 49: 1914).
- Mahendran, P. P. and Chandramani, P (1998). NPK-uptake, yield and starch content of potato cv. kufri Jyoti as influenced by certain biofertilizers. *Journal of the Indian Potato Association*, 27 (1-2): 50-52.
- Mahendran, P. P. and Kumar, N. (1998). Effect of biofertilizers on tuber yield and certain quality parameters of potato cv. kufri Jyoti. *South Indian Horticulture*, 46 (1-2): 47-48.

- Mahendran, P. P.; Kumar, N. and Saraswathy, S. (1996).** Studies on the effect of biofertilizers on potato. *South Indian Horticulture*, 44, (3-4): 79-82.
- Merghany, M. M. (1998).** Effect of irrigation systems and regimes in relation to farm yard manure levels on potato yield and quality in new reclaimed sandy soils. *Annals of Agric. Sci. Moshtohor*, 36 (2): 997-1014.
- Michel, K. A.; Gilles, J. K.; Ramilton, R. P. A. and Smith, F. (1956).** Colourimetric method for determination of sugars and related substances. *Anal. Chem.*, 28:3.
- Neweigy, N. A.; Ehsan A. Hanafy; Zaghoul, R. A. and El-Sayeda H. El-Badawy (1997).** Response of sorghum to inoculation with *Azospirillum*, organic and inorganic fertilization in the presence of phosphate solubilizing microorganisms. *Annals of Agric. Sci. Moshtohor*, 35 (3): 1383-1401.
- Page, A.L.; Miller, E. H. and Keeny, D. R. (1982).** *Methods of Soil Analysis*, part 2. 2nd Ed., Am. Soc. Agronomy, Inc. Madison, Wisconsin, U. S. A.
- Saad, O. A. O. and Hammad, A. M. M. (1998).** Fertilizing wheat plants with rock phosphate and VA-mycorrhizae as alternative for Ca-superphosphate. *Annals of Agric. Sci., Ain Shams Univ., Cairo*, 43 (2): 445-460.
- Saleh, E.A.; Nokhal, T. H.; El-Borollosy, M. A.; Fendrik, I.; Sharaf, M. S. and El-Sawy, M. (1998).** Effectiveness of dual inoculation with diazotrophs and vesicular arbuscular mycorrhizae on growth and medicinal compounds of *Datura stramonium*. *Arab. Univ. Journal Agric. Sci., Ain Sahms Univ., Cairo*, 6 (2): 343-355.
- Sharma, U. C. (1993).** Effect of farm yard manure on potato tuber yield and N uptake in Meghalaya. *Indian Journal of Hill Farming*, 3(2): 11-14.
- Shatokhina, S. F. and Khristenko, S. I. (1996).** Effect of different fertilizer system and seed inoculation on the microflora, biochemical and agrochemical characteristics of a typical chernozem, corn yield and quality of silage. *Eurasian Soil Science*, 24 (8): 456-462.
- Snedecor, G. W. and Cochran, W. G. (1989).** *Statistical Methods*, 8th Ed., Iowa State Univ., Press, Iowa, U. S. A.
- Sood, M. C. (1993).** Effect of tillage and mode of farm yard manure application on potato growth and yield at shimla. *Journal of the Indian Potato Association*, 22 (1-2): 83-85.
- Zaghoul, R. A.; Amer, A. A. and Mostafa, M. H. (1996).** Efficiency of some organic manures and bifertilization with *Azospirillum brasilense* for wheat manuring. *Annals of Agric. Sci., Moshtohor*, 34 (2): 627-640.
- Zahir, Z. A. and Muhammed, A. (1996).** Effectiveness of *Azotobacter* inoculation for improving potato yield under fertilized conditions. *Pakistan Journal of Agricultural sciences*, 33 (1-4): 1-5.
- Zahir, Z. A.; Muhammed, A.; Altaf, H.; Arshad, M.; Azam, M. and Hussain, A. (1997).** Effect of an auxin precursor tryptophane and *Azotobacter* inoculation on yield and chemical composition of potato under fertilized conditions. *Journal of Plant Nutrition*, 20 (6): 745-752.

فعالية التسميد الحيوي والعضوي على نمو وإنتاجية نباتات البطاطس

راشد عبد الفتاح زغول

قسم النبات الزراعي - كلية الزراعة بمشتهر - جامعة الزقازيق - فرع بنها

ملخص

كان الهدف من هذا البحث هو دراسة إمكانية استخدام الأسمدة الحيوية والتسميد العضوي كبديل للتسميد الكيماوي في إنتاج محصول البطاطس. وأقيمت لهذا الغرض تجربتان حقليتان بمزرعة التجارب بكلية الزراعة بمشتهر خلال موسمي ٢٠٠١، ٢٠٠٢. استُخدمت سلالتين فعاليتين من البكتريا المثبتة لأزوت الهواء الجوي هما *Azotobacter chroococcum*، *Azospirillum lipoferum* كذلك استُخدمت سلالة فعالة من البكتريا المذيبة للفوسفات وهي *B. megaterium var. phosphaticum*. تم استخدام سماد البيوجاز كسماد عضوي وسماد سلفات الأمونيوم كسماد كيماوي حيث أضيف سماد البيوجاز بمعدل ٦ طن/فدان (٩٠ كجم أزوت/فدان) كذلك تم استخدام نفس المعدل لوحدة الأزوت من سماد سلفات الأمونيوم ولقد أظهرت نتائج هذه الدراسة ما يلي:-

لوحظ عند التسميد بسماد البيوجاز أعلى نشاط للميكروبات ممثلاً في معدل ثاني أكسيد الكربون المنطلق في منطقة الريزوسفير. بينما كان أعلى نشاط لإنزيم النيتروجينيز عند تلقح تقاوي البطاطس عند الزراعة بمخلوط من الأزوتوباكتر والأزوسبيريللام. وعموماً فقد أوضحت النتائج أن معدل إطلاق ثاني أكسيد الكربون وكذلك نشاط إنزيم النيتروجينيز قد ازداد عند التلقيح بالبكتريا المذيبة للفوسفات مختلطة مع المعاملات المختلفة.

عند تلقح تقاوي البطاطس بالبكتريا المثبتة لأزوت الهواء الجوي (الأزوتوباكتر والأزوسبيريللام) مع إضافة نصف جرعة من سماد سلفات الأمونيوم أعطى ذلك قيمة أعلى من النيتروجين الأمونيومي والنترات في منطقة الريزوسفير مقارنة بالتسميد بجرعة كاملة من سماد سلفات الأمونيوم. أعطى سماد البيوجاز قيمة أعلى للنيتروجين والفوسفور الكلي في التربة مقارنة ببقية المعاملات. لوحظت فروقاً معنوية في صفات النمو التي درست حيث إزدادت هذه القياسات معنوياً عند التسميد بسماد البيوجاز وتلقيح التقاوي عند الزراعة بالبكتريا المذيبة للفوسفات. كذلك أدى التلقيح بالبكتريا المثبتة لأزوت الهواء الجوي مختلطة مع البكتريا المذيبة للفوسفات إلى زيادة معنوية في محتوى درنات البطاطس من الكربوهيدرات.

لم يشاهد فروق معنوية في محصول الدرنات/فدان عند التلقيح بالبكتريا المثبتة للأزوت والتسميد بسماد البيوجاز. وعموماً أوضحت هذه الدراسة أن محصول الدرنات/فدان في حالة التسميد الحيوي أو العضوي أعلى عنه في حالة التسميد الكيماوي. يمكن في ضوء نتائج هذه الدراسة أن يوصى باستخدام التسميد الحيوي أو العضوي في إنتاج محصول البطاطس كبديل للأسمدة الكيماوية خاصة وأنه يفضل حالياً استخدام هذه الأسمدة في الزراعة سواء كان ذلك للإنتاج المحلي أو للتصدير لأن معظم المستوردين يفضلون المنتج الزراعي الناتج من الزراعة العضوية والحيوية.